

Impairment of glutathione metabolism and its impact on other biochemical constituents in patients of diabetes mellitus

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Abstract: The intention of this study was to evaluate the levels of antioxidant enzymes glutathione peroxidase and glutathione reductase with their effect on lipid peroxidation and other biochemical constituents in patients of type 2 diabetes mellitus and compare with normal subjects of the same population. Amplified oxidative stress is extensively accepted contributor in the development and progression of diabetes and its complications. Usually diabetes is accompanied by greater production of reactive oxygen species (free radical) or impaired antioxidant defenses. NADPH dependent reduction of oxidized glutathione is catalyzed by the Glutathione reductase (GR) enzyme that serves to keep up intracellular glutathione supplies and a favorable redox status. We evaluate the status of glutathione reductase activity and other biochemical constituents in human serum of diabetic patients as well as in control subjects. This study was conducted on 40 Type 2 diabetic patients and 40 age and sex matched healthy control subjects. Fasting blood glucose (FBG), glycosylated hemoglobin (HbA_{1c}), Total Cholesterol, HDL, LDL, TG, glutathione peroxidase (GPX), glutathione reductase (GR) activity and lipid peroxidation product (MDA) were measured by using UV-visible spectrophotometric technique and compared with normal healthy persons. The results were statistically evaluated. The study revealed that FBG, HbA_{1c}, Total Cholesterol, LDL, TG and MDA were significantly higher whereas HDL, GPX and GR levels were significantly reduced in diabetics as compared to controls (P<0.05). Positive correlation of FBG with HbA_{1c} (r=0.529, P=0.0001), Total cholesterol with LDL (r=0.712, P=0.0001), GR with MDA (r=0.562, P=0.0001) whereas significant negative correlation was found between HbA_{1c} with GR (r=-.334, P=0.035), HbA_{1c} with MDA (r=-.340, P=0.032), HDL with TG (r=-.313, P=0.049), MDA with HbA_{1c} (r=-.340^{*}, P=0.032). It can be concluded that decreased levels of glutathione peroxidase and glutathione reductase along with high level of lipid peroxidation may be a useful markers of oxidative stress in type 2 diabetics. The increase in free radical activity in type 2 diabetes mellitus together with insulin resistance can lead to activation of stress-sensitive pathways, which play an important role in the complication of diabetes.

Keywords: Diabetes mellitus Type 2, glutathione peroxidase, glutathione reductase, lipid peroxidation product.

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INTRODUCTION

ROS are constantly formed in the human body and are removed by an antioxidant defense system¹. ROS are generally cytotoxic because they can cause damage to cellular components². The mechanism of free radical production include glucose autooxidation, protein glycation, advanced glycation end products formation, and activation of polyol pathway, ultimately resulting in oxidative stress in a variety of tissues³. Some biological parameters involved in cell defense against oxygen radicals are vitamin C and E, erythrocyte glutathione peroxidase, glutathione reductase, superoxide dismutase, and catalase. Tissue glutathione plays a central role in antioxidant defense. Unusually high levels of free radicals and the concurrent decline of antioxidant defense mechanisms can cause to damage cellular organelles and enzymes, greater lipid peroxidation, and insulin resistance progression. The consequences of oxidative stress can advance the development of complications of diabetes mellitus. Hyperglycemia also impairs the endogenous antioxidant defense system in many ways during diabetes⁴. An antioxidant is a molecule capable of slowing or preventing the oxidation of other molecules. Oxidation is a chemical reaction that transfers electrons from a substance to an oxidizing agent.

Oxidation reactions can produce free radicals, which start chain reactions that damage cells⁵. Antioxidants terminate these chain reactions by removing free radical intermediates, and inhibit other oxidation reactions by being oxidized themselves. As a result, antioxidants are often reducing agents such as thiols or polyphenols⁶.

Glutathione system is one of the chief defense mechanism against oxidative stress. Glutathione in reduced state detoxifies reactive oxygen species such as H₂O₂ and lipid peroxides directly or by a glutathione peroxidase (GPX) catalyzed reaction. Glutathione reductase (GR) catalyzes the NADPH dependent reduction of oxidized glutathione (GSS), help to keep up intracellular glutathione stores and a favorable redox state⁷. Determination of GSH/GSSG is a good measure of oxidative stress of an individual⁸. The chief defensive roles of glutathione against oxidative stress are that, it can act as several detoxifying enzyme's cofactor, scavenge hydroxyl radical and singlet oxygen directly, participate in amino acid transport across plasma membrane and redevelop Vitamins C and E back to their active forms⁹.

In this study, we want to evaluate the levels of an antioxidant enzyme glutathione peroxidase and reductase and their impact on other biochemical constituents including FBG, HbA_{1c}, HDL, LDL, TG,

and MDA in patients of type 2 diabetes and compared with normal subjects with good metabolic control, that whether the hyperglycemic induce variation in enzymatic levels affects other biochemical parameters or not in progression of diabetic complications.

MATERIALS AND METHODS

The study was conducted on 51 patients with type 2 diabetes both males and females (28 males and 23 females) between the age group 35-65 years who have registered at Baqai Institute of Diabetology and Endocrinology, Karachi, Pakistan. Fifty five (22 males and 33 females) age and sex matched control subjects were also selected from the general population at random for comparison. Ethical approval was obtained from the institutional review board (IRB) before the commencement of the study. Informed consent was taken from each individual at the time of recruitment in the study. All the patients who were diagnosed with type 2 diabetes using the ADA criteria i.e. FBG of ≥ 126 mg/dl were included in the study. The patients who had any recent clinical evidence of cardiac, renal or liver dysfunctions and any hemoglobinopathy were excluded from the study. Blood samples were collected in tubes with EDTA as anticoagulant. Plasma was separated and analyzed for other biochemical parameters such as FBG, HbA_{1c}, HDL, LDL, TG, GPX, GR activity and MDA. Fasting blood glucose was estimated by following glucose oxidase method on UV-visible spectrophotometer¹⁰.

The HbA_{1c} was analyzed by automatic D10 analyzer^{11, 12}. Total Cholesterol was estimated by enzymatic endpoint method (Randox Kit, Cat No: CH 200)¹³, HDL and LDL was estimated by CHOD-PAP Assay method (Randox kit, Cat No: CH 203)^{14,15}, Triglycerides was estimated by GPO-PAP method¹⁶. Glutathione Peroxidase estimated by Randox kit method (estimation based on principle of Paglia and Valentine)¹⁷. Glutathione reductase was estimated by Randox kit method^{18,19}. MDA was also estimated by Randox kit method on a UV-Visible Spectrophotometer^{20, 21}.

Data were statistically analyzed using Statistical Package for Social Sciences version 20 (SPSS Inc, Chicago, IL, USA). Independent samples were examined with student's t test. P-values and 95% confidence intervals (CI) were also calculated. P-value of < 0.05 was taken as significant for all comparisons.

RESULTS

The study was based 40 patients with diabetes type 2 (23 males and 17 females) and 40 healthy control subjects (17 males & 23 females). The Mean age of control was 45.17 ± 1.23 years and 50.87 ± 1.59 years for patients. The results of biochemical parameters (FBG, HbA_{1c}, Total Cholesterol, HDL, LDL, TG, GPX, GR and MDA) were compared in patients and control subjects. Significant increase in FBG, HbA_{1c}, Total Cholesterol, LDL, TG and MDA were found in diabetic patients as compared to healthy control subjects (Table 1). Pearson correlation was used to evaluate the impact of impaired glutathione metabolism on different biochemical parameters and to find the significant correlation between biochemical parameters in diabetic patients (Table 2). Positive correlations was found in FBG with HbA_{1c}, Total cholesterol with LDL, GR with MDA and negative correlation was found between HbA_{1c} with GR, HbA_{1c} with MDA, HDL with TG, (Table 2). The variation of biochemical constituents, antioxidant enzymes glutathione peroxidase & glutathione reductase and MDA level in control and diabetics on the basis of sex are shown in (Table 3).

Table 1: Comparison between control and diabetic subjects with respect to physical parameters and FBG, HbA_{1c}, lipid profile, GPX, GR and MDA levels.

Parameters	Control (n=40)	Diabetic Patients (n=40)	P-Value
Age (Years)	45.17±1.23	50.87±1.59	0.0058*
Ht (Cm)	164.71±1.70	166.19±1.64	0.5328
Wt (Kg)	60.80±1.73	69.72±2.07	0.0014*
BMI(Kg/m ²)	25.28±0.53	25.28±0.64	1.0000
FBG(mg/dl)	91.27±2.07	173.90±7.06	0.0001*
HbA _{1c} (%)	4.70±0.05	8.01±0.31	0.0001*
TC(mg/dl)	162.84±3.47	227.94±4.60	0.0001*
HDL(mg/dl)	89.22±5.38	34.02±1.74	0.0001*
LDL (mg/dl)	58.01±4.52	142.63±4.04	0.0001*
TG (mg/dl)	145.84±10.09	173.89±10.70	0.0602
GPX (mg/gHb)	47.05±3.00	37.76±3.18	0.0368
GR (U/L)	39.57±2.70	19.38±1.40	0.0001*
MDA(µM)	8.81±0.27	13.07±0.62	0.0001*

Ht(Height),Wt (Weight),BMI(Body mass index), Fasting blood glucose (FBG), Glycosylated hemoglobin (HbA_{1c}), Total Cholesterol (TC), High density lipoprotein (HDL), Low density lipoprotein (LDL), Triglyceride (TG), Glutathione peroxidase (GPX), Glutathione Reductase (GR), Malondialdehyde(MDA), n = no of subjects, values are represented as mean ± SEM (Standard error of mean), * P < 0.05 is considered to be statistically Significant.

DISCUSSION

The mechanism of free radical production include glucose autoxidation, protein glycation, advanced glycation end products formation, and

activation of polyol pathway, ultimately resulting in oxidative stress in a variety of tissues²².

Our study concluded that the levels of all the biochemical constituents such as Blood Glucose, Glycosylated hemoglobin, Total Cholesterol, LDL and Triglycerides were higher in diabetic patients as compare to control subjects which shows that the disturbance in carbohydrate metabolism affects all the other metabolic pathways. While the levels of antioxidant enzymes Glutathione peroxidase and reductase are lower in type 2 diabetic patients as compare to control subjects which shows that hyperglycemia induce oxidative stress which may occur through depletion of NADPH and consequently disturbance of glutathione metabolism²³.

Table 2: Correlations.

	Glucose	HbA1c	TC	HDL	LDL	TG	GPX	GR	MDA
Glucose	1	.529**	-.179	.141	-.050	.051	-.048	-.052	.496**
		.000	.268	.387	.759	.756	.769	.752	.001
	40	40	40	40	40	40	40	40	40
HbA1c	.529**	1	-.236	.289	-.066	-.082	-.109	-.334*	.060
	.000		.143	.070	.684	.613	.503	.035	.713
	40	40	40	40	40	40	40	40	40
TC	-.179	-.236	1	-.288	.712**	.170	.159	.186	.284
	.268	.143		.071	.000	.295	.327	.251	.076
	40	40	40	40	40	40	40	40	40
HDL	.141	.289	-.288	1	-.088	-.313*	-.233	-.051	-.015
	.387	.070	.071		.588	.049	.148	.756	.925
	40	40	40	40	40	40	40	40	40
LDL	-.050	-.066	.712**	-.088	1	-.115	.063	.252	.293
	.759	.684	.000	.588		.479	.700	.117	.067
	40	40	40	40	40	40	40	40	40
TG	.051	-.082	.170	-.313*	-.115	1	.033	.052	-.147
	.756	.613	.295	.049	.479		.838	.749	.367
	40	40	40	40	40	40	40	40	40
GPX	-.048	-.109	.159	-.233	.063	.033	1	.022	.037
	.769	.503	.327	.148	.700	.838		.893	.819
	40	40	40	40	40	40	40	40	40
GR	-.052	-.334*	.186	-.051	.252	.052	.022	1	.270
	.752	.035	.251	.756	.117	.749	.893		.092
	40	40	40	40	40	40	40	40	40
MDA	.496**	.060	.284	-.015	.293	-.147	.037	.270	1
	.001	.713	.076	.925	.067	.367	.819	.092	
	40	40	40	40	40	40	40	40	40

Depletion of NADPH occurs as a result of activation of polyol pathway in hyperglycemic condition which is also an NADPH dependent pathway²⁴. Inconsistency was observed in biomarkers for oxidative stress such as glutathione peroxidase, glutathione reductase in diabetics. Decreased levels of glutathione and elevated concentrations of MDA are also observed in diabetes²⁵.

In present study significant decrease in the activities of antioxidant enzymes Glutathione Peroxidase and Glutathione Reductase are found between diabetic and non-diabetic individuals and this finding is supported by many studies^{26, 27}. Blood GSH was significantly decreased in different phases

of type2 DM such as: glucose intolerance and early hyperglycemia, within two years of diagnosis and before development of complications and in poor glycemic control. The pathophysiological significance of decreased glutathione levels in DM remains to be unclear. Some studies revealed no difference in whole blood GR activity in type1 and type2 DM patients compared to control subjects²⁸.

Current study shows positive correlation between Fasting Blood Glucose level with HbA1c & MDA and negative correlation with enzymatic levels (GPX & GR) (Figure 1). On the other hand negative correlation was found between HbA1c level and enzymatic levels (GPX & GR) and positive correlation between HbA1c and MDA(Figure 2). The variation of Biochemical constituents, antioxidant enzymes glutathione peroxidase and glutathione reductase and MDA level in control and diabetics on the basis of sex are shown in (Table 3).On the basis of these finding it is suggested that antioxidant enzymes levels should be utilized in clinical practice to improve vascular risk prediction in diabetics.

Table 3: Variations of FBG, HbA1c, lipid profile, GPX, GR and MDA in control and diabetic male and female subjects.

Parameters	Control (n=40)		Diabetic patients (n=40)	
	Male (n= 17)	Female (n=23)	Male (n= 23)	Female (n=17)
FBG	86.34 ± 2.79	94.91 ±2.77	178.30 ±9.83*	167.94 ±10.13*
HbA1c	4.57 ± 0.07	4.79 ±0.08	8.30 ±0.39*	7.62 ±0.52*
TC	166.72 ±4.11	159.98 ±5.21	228.85 ±5.81*	226.71 ±7.64*
HDL	92.89 ±9.26	86.50 ±6.51	33.87 ±2.06*	34.22 ±3.07*
LDL	49.80 ±6.22	64.08 ±6.19	141.00 ±5.50*	144.84 ±6.09*
TG	167.23 ±17.35	130.02 ±11.19	182.60 ±15.27	162.1 ±14.44
GPX	49.85 ±5.25	44.98 ±3.53	33.71 ±3.93*	43.22 ±5.11
GR	41.03 ±4.00	38.50 ±3.72	18.85 ±2.06*	20.10 ±1.83*
MDA	8.37± 0.29	9.13 ±0.42	13.70 ±0.90*	12.21 ±0.78

Fasting blood glucose (FBG), Glycosylated hemoglobin (HbA_{1c}), Total Cholesterol (TC), High density lipoprotein (HDL), Low density lipoprotein (LDL), Triglyceride (TG), Glutathione peroxidase (GPX), Glutathione Reductase (GR), Malondialdehyde (MDA), n = no: of subjects, values are represented as mean ± S.E.M (Standard error of mean). * Statistically significant as compared to control.

Decreased activity and efficiency of cellular antioxidant mechanisms with concurrent increased lipid peroxidation represent the pathogenic

connection between hyperglycemia and expansion of endothelial dysfunction. Furthermore, the extent of oxidative stress and lack of defensive antioxidant mechanisms in Type 2 Diabetic patients is reliant on the metabolic control of diabetes and the incidence of complications. Intensity of oxidative stress in Type 2 diabetic patients is greater when compared with normal healthy individuals as controls. Antioxidant enzymes (Glutathione Peroxidase and Glutathione Reductase) levels as surrogate marker in early detection of diabetic complications. Suggesting it that, they may contribute in finding micro and macrovascular complications in diabetics.

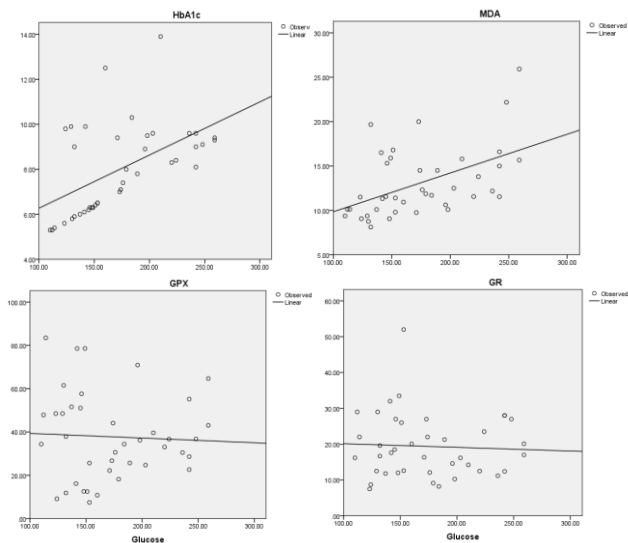


Figure 1: Correlation between FBG level with HbA1c and MDA and negative correlation with enzymatic levels (GPX and GR).

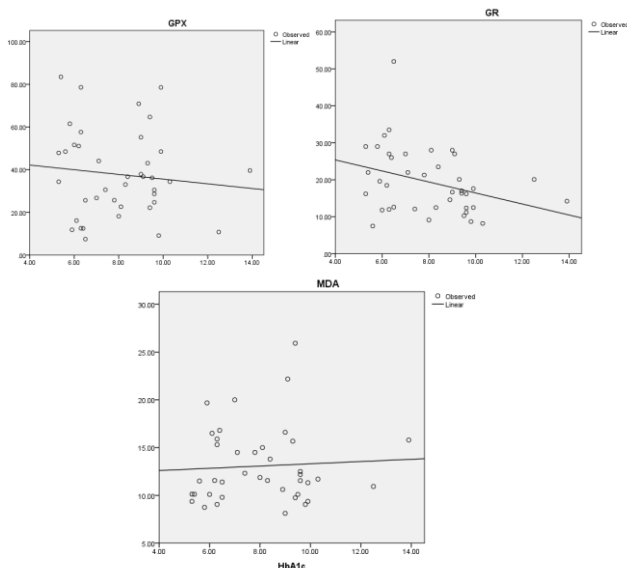


Figure 2: Correlation between HbA1c level with enzymatic levels (GPX and GR) and positive correlation between HbA1c and MDA.

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REFERENCES

- Halliwel B. The role of oxygen radicals in human disease, with particular reference to the vascular system. *Haemostasis*. 1993; 23:118-126.
- Bae YS, Kang SW, Seo MS, Baines IC, Tekle E, Chock PB and Rhee SG: Epidermal growth factor (EGF)-induced generation of hydrogen peroxide. Role in EGF receptor-mediated tyrosine phosphorylation. *J. Biol. Chem.*, 1997; 272: 217-221.
- Baynes JW and Thorpe SR. Role of Oxidative Stress in Diabetic Complications: a new perspective on an old paradigm. *Diabetes* 1999; 48: 1-9.
- Laight DW, Carrier MJ and Anggard EE. Antioxidants, diabetes and endothelial dysfunction. *Cardiovasc. Res.*, 2000; 47: 457-464.
- Sies Helmut. Oxidative stress: Oxidants and antioxidants. *Exp. Physiol.*, 1997; 82: 291-295.
- Valko M, Leibfritz D, Moncol J, Cronin M, Mazur M and Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. and Cell Biol.*, 2007; 39: 44-84.
- Rahman K. Studies on free radicals, antioxidants, and co-factors. *Clin. Interv. Aging*. 2007; 2: 219-236.
- Dröge W. Aging-related changes in the thiol/disulfide redox state: implications for the use of thiol antioxidants. *Exper. Geront.*, 2002; 37: 1333-1345.
- Masella R, Di Benedetto R and Vari R. Novel mechanisms of natural antioxidant compounds in biological systems: involvement of glutathione and glutathione-related enzymes. *J. Nutr. Biochem.*, 2005; 16:577-586.
- Tietz. *Textbook of Clinical Chemistry and Molecular Diagnostics*, Fourth Edition. Carl A. Burtis, Edward R. Ashwood, and David E. Bruns, editors. St. Louis, MO: Elsevier Saunders, 2006.
- Yasmeen F, Mumtaz A, Saleem-Uz-Zaman A and Qureshi SA. Comparison of cation exchange HPLC and Immunoturbidimetric method for determination of HbA_{1c}. *Biomedica*, 2011; 27: 161-165.
- Marzullo C and Minery M. Evaluation of D10 hemoglobin testing system for hemoglobin A1C assay. *Ann. Biol. Clin.*, 2008; 66: 95-99.
- Roeschlau P, Bernt E and Gruber W. Enzymatic determination of total cholesterol in serum. *Z. Klin. Chem. Klin. Biochem.*, 1974; 12: 403-407.
- Rifai N, Bachorik PS and Albers JJ. Lipids, lipoproteins and apolipoprotein. In: Burtis CA, Ashwood ER, editors. *Tietz Textbook of clinical chemistry*. 3rd edition. Philadelphia: WB Saunders Company. 1999; 809-61.
- Harald T. Specimen Collection and Handling: Standardization of blood sample collection. *Methods mol. Biol.*, 2008; 428: 35-42.
- Jacobs NJ and VanDenmark PJ. Enzymatic determination of serum triglyceride. *Biochem. Biophys.*, 1960; 88: 250-255.

17. Paglia DE and Valentine WN. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *J. Lab Clin. Med.*, 1967; 70:158-169.
18. Goldberg DM and Spooner RJ. Glutathione reductase. Bergmeyer HU eds. *Methods of enzymatic analysis* 3rd ed. 1983: pp 258-265.
19. Melissinos KG, Delidov AZ, Varsov AG, Begjetti SS and Drivas GJ. Determination of glutathione reductase activity. *Nephron*, 1981; 28: 76-79.
20. Esterbauer H, Schaur RJ and Zollner H. Chemistry and biochemistry of hydroxynonenal, malondialdehyde and related aldehydes. *Free Radical Biol. Med.*, 1991; 11: 81-128.
21. Gerard-Monnier D, Erdelmeier I, Regnard K, Mozehenry N, Yadan JC and Chaudierel J. Reactions of 1-Methyl-2-phenylindole with malondialdehyde and 4-hydroxyalkenals Analytical applications to a colorimetric assay of lipid peroxidation. *Chem. Res. Toxicol.*, 1998; 11: 1176–1183.
22. Imlay JA. Pathways of oxidative damage. *Ann. Rev. Microbiol.*, 2003; 57: 395–418.
23. Ferdinando G and Michael B. Oxidative stress and diabetic complications. *Circ. Res.*, 2010; 107: 1058-1070.
24. Chung SS, Ho EC, Lam KS and Chung SK. Contribution of polyol pathway to diabetes-induced oxidative stress. *J. Am. Soc. Nephrol.*, 2003; 14: S233–S236.
25. Ozdemir G, Ozden M, Maral H, Kuskay S, Cetinalp P and Tarkun I. Malondialdehyde, glutathione, glutathione peroxidase and homocysteine levels in type 2 diabetic patients with and without microalbuminuria. *Ann. Clin. Biochem.*, 2005; 42: 99-104.
26. Thornalley PJ, McLellan AC, Lo TW, Benn J and Sönksen PH. Negative association between erythrocyte reduced glutathione concentration and diabetic complications. *Clin. Sci.*, 1996; 91: 575–582.
27. Yoshida K, Hirokawa J, Tagami S, Kawakami Y, Urata Y and Kondo T. Weakened cellular scavenging activity against oxidative stress in diabetes mellitus: regulation of glutathione synthesis and efflux *Diabetologia*. 1995; 38: 201-210.
28. Vijayalingam S, Parthiban A, Shanmugasundaram KR and Mohan V. Abnormal antioxidant status in impaired glucose tolerance and non-insulin-dependent diabetes mellitus. *Diab. Med.*, 1996; 13: 715-719.